



# Prdx1 Inhibits Tumorigenesis via Regulating PTEN/AKT Activity

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# Prdx1 inhibits tumorigenesis via regulating PTEN/AKT activity

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It is widely accepted that reactive oxygen species (ROS) promote tumorigenesis. However, the exact mechanisms are still unclear. As mice lacking the peroxidase peroxiredoxin1 (Prdx1) produce more cellular ROS and die prematurely of cancer, they offer an ideal model system to study ROS-induced tumorigenesis. Prdx1 ablation increased the susceptibility to Ras-induced breast cancer. We, therefore, investigated the role of Prdx1 in regulating oncogenic Ras effector pathways. We found Akt hyperactive in fibroblasts and mammary epithelial cells lacking Prdx1. Investigating the nature of such elevated Akt activation established a novel role for Prdx1 as a safeguard for the lipid phosphatase activity of PTEN, which is essential for its tumour suppressive function. We found binding of the peroxidase Prdx1 to PTEN essential for protecting PTEN from oxidation-induced inactivation. Along those lines, Prdx1 tumour suppression of Ras- or ErbB-2-induced transformation was mediated mainly via PTEN.

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## Introduction

Peroxiredoxins (Prdxs) are a superfamily of small non-seleno peroxidases (22–27 kDa) currently known to possess six

mammalian isoforms. Although their individual roles in cellular redox regulation and antioxidant protection are quite distinct, they all catalyse peroxide reduction to balance cellular H<sub>2</sub>O<sub>2</sub> levels essential for signalling and metabolism (Rhee, 2006). Prdxs I–IV share the common CxxC motif and use thioredoxin (Trx) as an electron donor (Chae *et al*, 1994a). On reaction with peroxides, the Prdxs' 'peroxidatic' cysteine (Cys51 in Prdx1) is oxidized into a sulfenic acid intermediate, which then forms a disulfide bond with the 'resolving' cysteine (Cys172 in Prdx1) from the other homodimer's subunit. Catalytic reduction of peroxides results in different oxidation states of the catalytic cysteine. Lower oxidation states (sulfenic acid) are reduced back to thiol by the Trx system. Higher oxidation states such as sulfinic acid can be reduced back to thiol by sulfiredoxin, whereas the oxidation to sulfonic acid is irreversible and results in Prdx degradation (Neumann and Fang, 2007). Loss of Prdx1 shortens the life span of mice due to the development of hemolytic anemia and cancer. Analysis of cells and mice lacking Prdx1 suggested that it not only regulated H<sub>2</sub>O<sub>2</sub> levels, but also that it possessed the properties of a tumour suppressor (Neumann *et al*, 2003), which was further supported by the finding that Prdx1 interacts with c-Myc thereby selectively inhibiting c-Myc transcriptional activity (Egler *et al*, 2005).

Peroxides are known to modify protein tyrosine phosphatases (PTPs) by oxidation. PTP catalytic property depends on a thiolate anion of a low pK<sub>a</sub> cysteine residue (pK<sub>a</sub> 4.7–5.4) located in the conserved motif of its active site (Zhang and Dixon, 1993; Peters *et al*, 1998). This highly nucleophilic group makes the initial attack on the phosphate group of the substrate, but renders the PTP extremely susceptible to oxidation resulting in its inactivation. The PTP and tumour suppressor PTEN exhibits phosphatase activity towards phosphoinositides and tyrosine phosphates and is known to be inactivated through H<sub>2</sub>O<sub>2</sub>-mediated oxidation or through growth factor signalling, including epithelial growth factor (EGF) or platelet-derived growth factor (PDGF) (Leslie *et al*, 2003; Kwon *et al*, 2004). Analysis of human recombinant PTEN revealed that two of the five cysteines in its N-terminal phosphatase domain (PTD) (Cys71 and Cys124) form a disulfide bond after oxidation, which resulted in the transient inhibition of its phosphatase activity and allowed oxidized PTEN to migrate faster on a non-reducing SDS-PAGE (Lee *et al*, 2002; Kwon *et al*, 2004). In stimulated macrophages, PTEN oxidation led to the temporary inhibition of its phosphatase activity and in the phosphorylation of Akt on Serine 473 (Leslie *et al*, 2003).

PTEN deficiency is a hallmark of many human tumours (Keniry and Parsons, 2008) and is accompanied by enhanced cell proliferation, decreased cell apoptosis and increased Akt activity (Stambolic *et al*, 1998). Akt is regulated by PI3K and PTEN, thus playing a central role particularly in Ras- and

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ErbB-2-induced transformation. Akt activity has been proven crucial for the initiation of this transformation, both *in vitro* and *in vivo* (Sheng *et al*, 2001; Hutchinson *et al*, 2004; Lim and Counter, 2005) and phosphatase inactive PTEN (PTEN Cys124Ser) is incapable of suppressing Ras-induced transformation either *in vitro* or *in vivo* (Tolkacheva and Chan, 2000; Koul *et al*, 2002).

Although it is known that PTEN activity is negatively regulated through oxidation, the existence of a protective mechanism inhibiting such inactivation has not been proposed. On the basis of our findings, we propose here that Prdx1 promotes PTEN tumour suppressive function by binding PTEN and protecting its lipid phosphatase activity from H<sub>2</sub>O<sub>2</sub>-induced inactivation. This way, Prdx1 controls excessive cellular Akt activity and reduces the susceptibility to H-Ras and ErbB-2-induced transformation.

## Results

### Prdx1 inhibits Ras-induced transformation through its peroxidase activity

We tested first Prdx1 peroxidase activity under physiological conditions and measured endogenous H<sub>2</sub>O<sub>2</sub> release in growing cells over time. Extracellular H<sub>2</sub>O<sub>2</sub> buildup from Prdx1<sup>-/-</sup>;Prdx1<sup>WT</sup>MEFs was less compared with Prdx1<sup>-/-</sup>MEFs during the 240 min measured. In contrast, Prdx1<sup>-/-</sup>;Prdx1<sup>C51/172S</sup>MEFs resulted in an excess of extracellular H<sub>2</sub>O<sub>2</sub> buildup (Supplementary Figure S1A). Rates calculated for extracellular H<sub>2</sub>O<sub>2</sub> buildup (pmol/min) confirmed that Prdx1<sup>-/-</sup>;Prdx1<sup>WT</sup>MEFs released H<sub>2</sub>O<sub>2</sub> at a 30% lower rate than Prdx1<sup>-/-</sup>MEFs, whereas Prdx1<sup>-/-</sup>;Cys51/172SMEFs released H<sub>2</sub>O<sub>2</sub>-release 1.5-fold more than Prdx1<sup>-/-</sup>MEFs (Figure 1A, left panel). Western blotting analysis further confirmed that Prdx1 was functioning as a peroxidase, as Prdx1<sup>WT</sup> protein formed mostly DTT-reducible (data not shown) H<sub>2</sub>O<sub>2</sub>-scavenging dimeric structures (Figure 1A, right panel), whereas Prdx1<sup>Cys51/172S</sup> did not, as dimer formation depends on Cys51 and Cys172 disulfides (Chae *et al*, 1994b). The remaining Prdx1 monomers were not over-oxidized and therefore active (unlike Prdx1 monomers in H<sub>2</sub>O<sub>2</sub>-treated Prdx1<sup>-/-</sup>;Prdx1<sup>WT</sup>MEFs). Prdx1 peroxidase activity was required to suppress H-Ras-induced tumour suppression, as Prdx1<sup>-/-</sup>MEFs transduced with retrovirus expressing H-Ras<sup>V12</sup> formed two-fold more colonies in soft agar compared with Prdx1<sup>+/+</sup>;H-Ras<sup>V12</sup>MEFs (Figure 1B, left panel). Expression of exogenous Prdx1<sup>WT</sup> reduced colony formation by 50% in Prdx1<sup>-/-</sup>;H-Ras<sup>V12</sup>MEFs and by 20% in Prdx1<sup>+/+</sup>;H-Ras<sup>V12</sup>MEFs, whereas exogenously expressed Prdx1<sup>C51/172S</sup> modestly increased colony formation in Prdx1<sup>-/-</sup>;H-Ras<sup>V12</sup>MEFs and Prdx1<sup>+/+</sup>;H-Ras<sup>V12</sup>MEFs. Western blot analysis ensured that re-expression of Prdx1 proteins in Prdx1<sup>-/-</sup>MEFs did not exceed those Prdx1 levels found in Prdx1<sup>+/+</sup>MEFs (Figure 1B, right panel). Prdx1<sup>-/-</sup>;H-Ras<sup>V12</sup>MEFs appeared also more spindle-like than Prdx1<sup>+/+</sup>;H-Ras<sup>V12</sup>MEFs, whereas expression of Prdx1<sup>WT</sup> in Prdx1<sup>-/-</sup>;H-Ras<sup>V12</sup>MEFs reversed the spindle-like morphology. Expression of exogenous Prdx1<sup>C51/172S</sup>, however, promoted spindle-like cell morphology in both Prdx1<sup>-/-</sup>;H-Ras<sup>V12</sup>MEFs and Prdx1<sup>+/+</sup>;H-Ras<sup>V12</sup>MEFs (Supplementary Figure S1B). Analysis of the Prdx1 peroxidase activity in H-Ras<sup>V12</sup>-transformed MEFs showed that Prdx1<sup>-/-</sup>;Prdx1<sup>WT</sup>;H-Ras<sup>V12</sup>MEFs released H<sub>2</sub>O<sub>2</sub> at a 26%

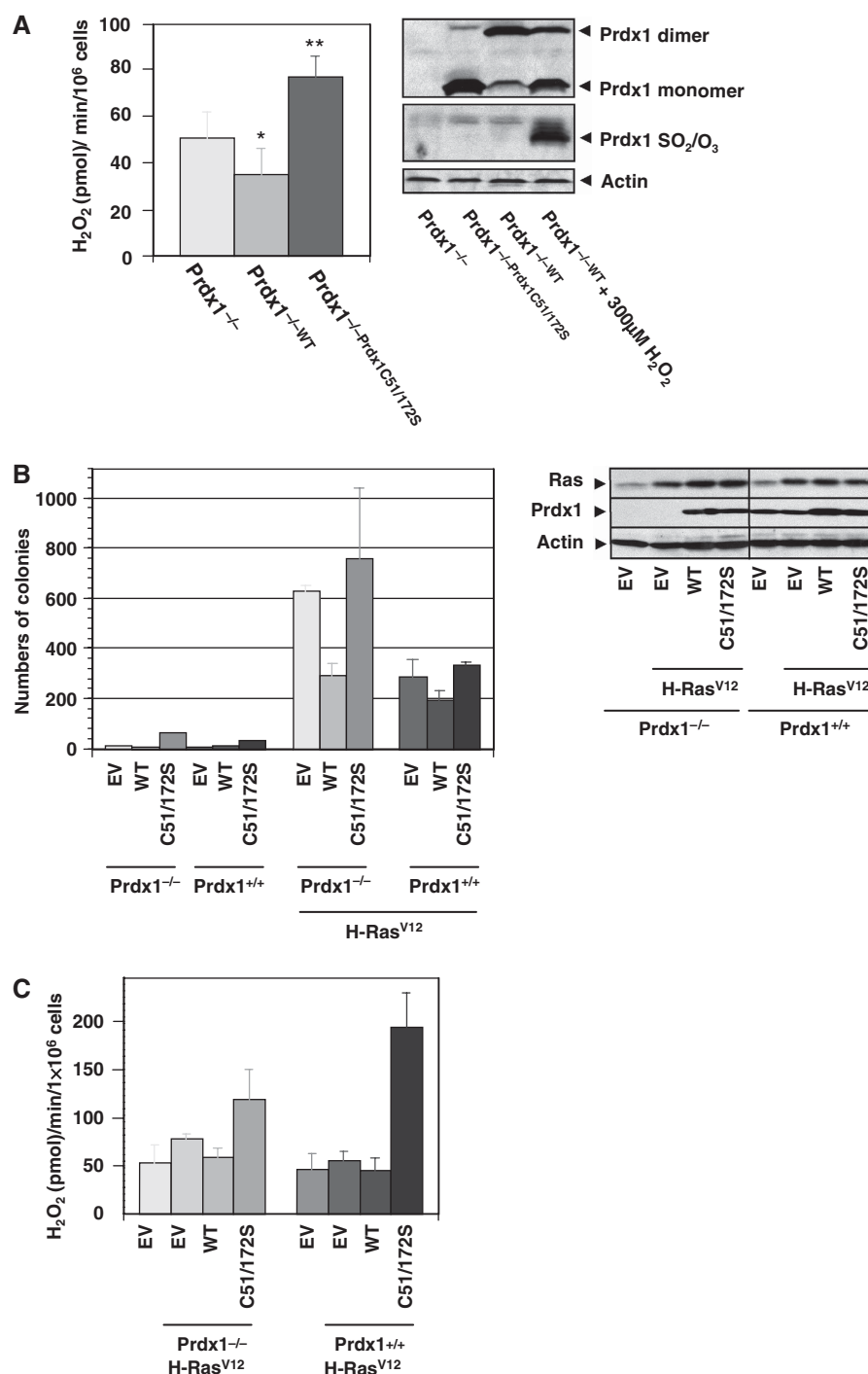
lesser rate than Prdx1<sup>-/-</sup>;H-Ras<sup>V12</sup>MEFs, which was comparable to the rate found Prdx1<sup>-/-</sup>EVMEFs. In contrast, Prdx1<sup>-/-</sup>;C51/172S;H-Ras<sup>V12</sup>MEFs had a 1.5-fold increased rate of H<sub>2</sub>O<sub>2</sub> release compared with Prdx1<sup>-/-</sup>;Prdx1<sup>WT</sup>;H-Ras<sup>V12</sup>MEFs (Figure 1C).

### Loss of Prdx1 promotes PTEN oxidation and Akt activation

We investigated now whether Prdx1 regulates H-Ras effector pathways known to promote transformation, including PI3K/Akt, RalGEF or MAPK pathways (Li *et al*, 2004; Lim and Counter, 2005). Treatment of MEFs with H<sub>2</sub>O<sub>2</sub> (50 μM) for increasing periods of time enhanced Akt phosphorylation on Ser473 (pAkt<sup>Ser473</sup>) and Thr308 (pAkt<sup>Thr308</sup>) at a higher level in Prdx1<sup>-/-</sup>MEFs compared with Prdx1<sup>+/+</sup>MEFs (Figure 2A). This finding correlated with a steady increase of PTEN oxidation over time in Prdx1<sup>-/-</sup>MEFs, whereas in Prdx1<sup>+/+</sup>MEFs PTEN oxidation plateaued after 6–8 min (Figure 2B and C). Phosphorylation of Akt on serine 473 (pAkt<sup>Ser473</sup>) and PTEN oxidation increased also in Prdx1<sup>-/-</sup>MEFs compared with Prdx1<sup>+/+</sup>MEFs when exposed to increasing amounts of H<sub>2</sub>O<sub>2</sub> (Supplementary Figure S2A and B). Similar data were obtained by inducing H<sub>2</sub>O<sub>2</sub> endogenously through treating cells with PDGF, as it led to a larger increase in (1) pAkt<sup>Ser473</sup> and pAkt<sup>Thr308</sup> (Figure 2D) and (2) PTEN oxidation in Prdx1<sup>-/-</sup>MEFs when compared with Prdx1<sup>+/+</sup>MEFs (Figure 2E). Although PTEN oxidation steadily increased in Prdx1<sup>-/-</sup>MEFs, it decreased in Prdx1<sup>+/+</sup>MEFs over time (Figure 2E and F). Levels of pAkt<sup>Ser473</sup> and PTEN oxidation was also increased in Prdx1<sup>-/-</sup>MEFs compared with Prdx1<sup>+/+</sup>MEFs when exposed to increasing amounts of H<sub>2</sub>O<sub>2</sub> or PDGF (Supplementary Figure S2A–D). Lastly, we confirmed that lack of Prdx1 enhanced basal and H<sub>2</sub>O<sub>2</sub>-induced phosphorylation of Akt substrates in Prdx1<sup>-/-</sup>MEFs more than in Prdx1<sup>+/+</sup>MEFs (Figure 2G).

### Prdx1 interacts with PTEN

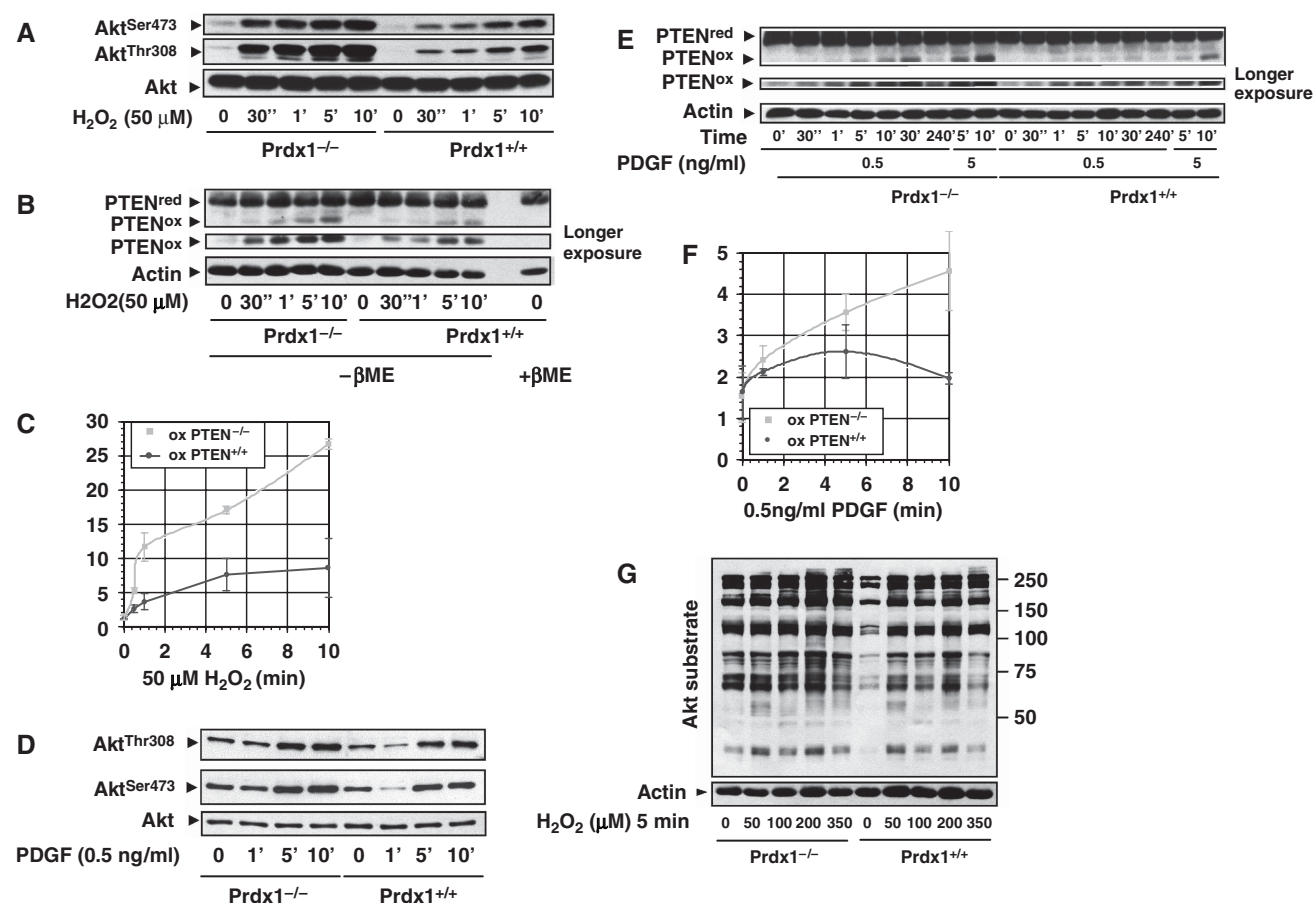
PTEN forms a complex with endogenous and epitope-tagged HA-Prdx1 (Figure 3A). This complex was regulated by oxidative stress, as co-expressed epitope-tagged Myc-PTEN and HA-Prdx1 dissociated under increasing concentrations of oxidative stress (Figure 3B, upper panel). In contrast, increasing dosages of H<sub>2</sub>O<sub>2</sub> did not disrupt binding of Prdx1<sup>Cys51/172S</sup> with PTEN (Figure 3C). Immunoblotting confirmed equal expression of proteins (Supplementary Figure S3A and B). Peroxidatic Cys51 may regulate the H<sub>2</sub>O<sub>2</sub>-induced Prdx1:PTEN complex disruption as Prdx1<sup>C51S</sup> and Prdx1<sup>C51/172S</sup> proteins bind more to wild-type PTEN than Prdx1<sup>C172S</sup> (Figure 3D). We further confirmed a physical association between Prdx1 and PTEN by using GST-pull down of GST-Prdx1 and His-PTEN (Figure 3E). Mutational analysis suggested that Prdx1 interacts within the C2 domain of PTEN (amino acids 186–274) (Figure 3F; Supplementary Figure S3C and D) and PTEN with the N terminus of Prdx1 (amino acids 1–40) and the C terminus of Prdx1 (amino acids 157–199) (Figure 3F, lower schematic and Supplementary Figure S3G). Computational analysis allowed us to narrow those interaction surfaces (Figure 3G). The images of Prdx1 and PTEN were generated from coordinates from the protein database 1d5r (Lee *et al*, 1999) and 2z9s (Matsumura *et al*, 2008), respectively. As shown for PTEN, the region of 186–251 consists of two double-stranded anti-parallel sheets con-



**Figure 1** (A) Left panel: rate of  $\text{H}_2\text{O}_2$  release per minute was analysed by calculating the linear slope of data obtained in Supplementary Figure S1A.  $\text{H}_2\text{O}_2$  release is expressed per  $1 \times 10^6$  cells per minute. *P*-values were calculated using an unpaired Student's *t*-test. Rate of  $\text{H}_2\text{O}_2$  release in *Prdx1*<sup>-/-</sup>MEFs compared with either *Prdx1*<sup>-/-</sup>Prdx1<sup>WT</sup> MEFs (\**P* = 0.04) or *Prdx1*<sup>-/-</sup>Prdx1<sup>C51/172S</sup> (\*\**P* = 0.001). Right panel: protein levels and oxidation status of expressed Prdx1 in MEFs used in left panel. Protein lysates were prepared as described under Materials and methods and analysed under non-reducing conditions. Status of over-oxidized Prdx1 was tested by using an antibody recognizing Prdx1 (Prdx1-SO<sub>2</sub>/SO<sub>3</sub>) (Woo *et al.*, 2003). (B) Left panel: *Prdx1*<sup>-/-</sup>MEFs and *Prdx1*<sup>+/+</sup>MEFs were infected with retroviral constructs carrying genes for *Prdx1*<sup>WT</sup> or *Prdx1*<sup>C51/172S</sup>. MEFs were further infected with retrovirus expressing either H-Ras<sup>V12</sup> or puromycin resistance gene only (EV) and plated in duplicates in soft agar. Colonies were counted after 18–20 days. Right panel: proteins of clones used in soft agar experiment were analysed for expression levels of Prdx1 and Ras. (C) Rate of  $\text{H}_2\text{O}_2$  release per minute was analysed by calculating the linear slope of data obtained in MEFs used in (B, left panel). For each clone, six wells were plated and analysed. The experiment shown here is representative of three independent studies from two different sets of MEF clones obtained from *Prdx1* littermates. *P*-values were calculated using an unpaired Student's *t*-test. \**P* = 0.045; \*\**P* = 0.004. Differences in *Prdx1*<sup>+/+</sup>MEFs were not statistically significant.

nected through two flexible loops. Similarly, the 1–40 region of Prdx1 consists of a single double-stranded anti-parallel sheet and another, surface exposed single sheet strand. In

Figure 3G, the Prdx-1 protein is shown only as the monomer and the helical region that lies at the dimer interface, residues 183–199, are also positioned in a surface exposed position



**Figure 2** (A) Prdx1<sup>-/-</sup> MEFs and Prdx1<sup>+/+</sup> MEFs were stimulated with H<sub>2</sub>O<sub>2</sub> as indicated. Protein lysates were collected under argonized conditions by scraping cells into argon-purged lysis buffer (see Materials and methods) and analysed under non-reducing conditions on SDS-PAGE. Akt phosphorylation was detected on Ser473 and Thr308. Akt protein as loading control. (B) Prdx1<sup>-/-</sup> MEFs and Prdx1<sup>+/+</sup> MEFs protein lysates were treated as described under (A) and analysed for oxidized PTEN proteins. Actin as loading control. (C) Levels of oxidized PTEN proteins were evaluated by quantifying oxidized and reduced PTEN using a Fuji imaging system (LAS 3000) and related software (ImageGauge). Quantifications of staining intensities were obtained by analysing protein bands from the same ECL obtained film exposure. The y-axis represents staining intensities in arbitrary units of oxidized PTEN. Curves represent data from three independent experiments. (D) Prdx1<sup>-/-</sup> MEFs and Prdx1<sup>+/+</sup> MEFs were stimulated with PDGF as indicated. Protein lysates were collected and analysed as described under (A). (E) Prdx1<sup>-/-</sup> MEFs and Prdx1<sup>+/+</sup> MEFs protein lysates were treated as described under (D) and analysed for oxidized PTEN proteins. Actin as loading control. (F) Data analysis was done as described under (C). All experiments shown are representative of at least three independent studies including two sets of MEF clones from Prdx1 littermates. (G) Serum starved MEFs were stimulated with H<sub>2</sub>O<sub>2</sub> as indicated. Protein lysates were collected and analysed as described under (A). Akt substrates were detected by western blotting using an phospho-Akt substrate antibody (RXRXXS/T).

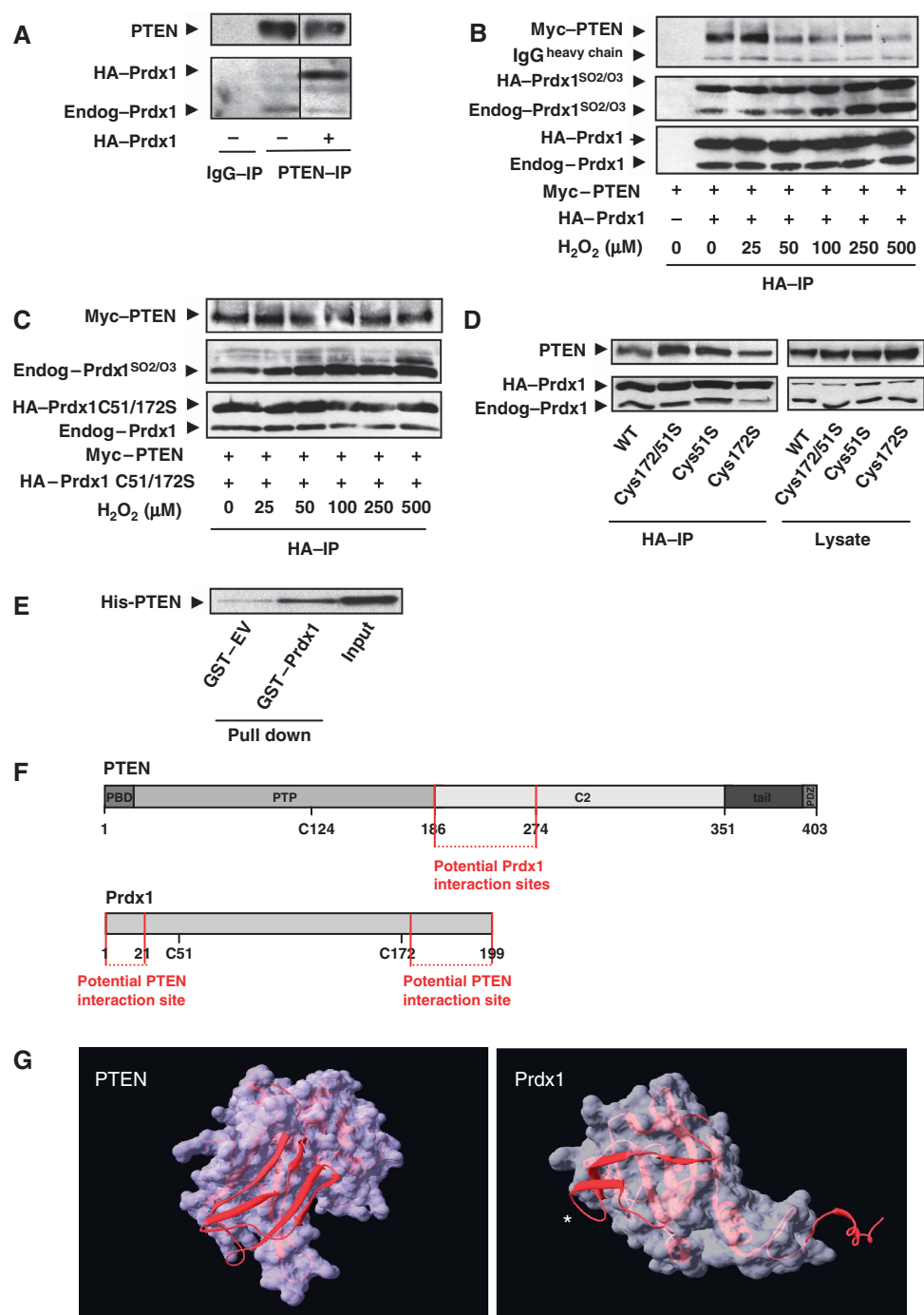
that is obviously well poised to mediate a protein-protein interaction. Deleting the identified interaction sites in PTEN and Prdx1 altered enzyme activity of both enzymes (data not shown), which did not allow us to study the role of a physical interaction in protecting PTEN lipid phosphatase activity.

#### **Prdx1 interaction with PTEN protects and promotes PTEN lipid phosphatase activity under oxidative stress**

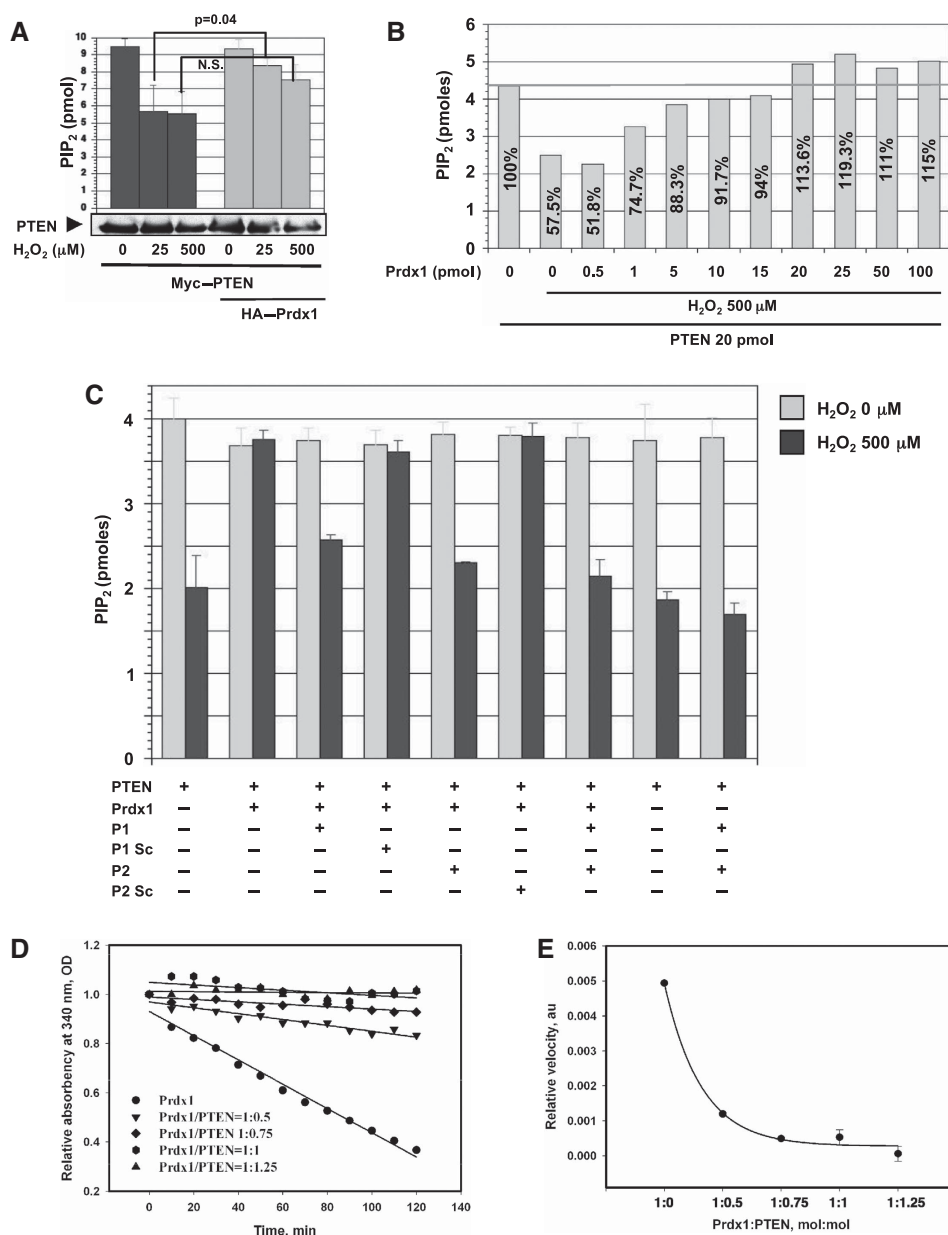
PTEN lipid phosphatase activity was fully protected by Prdx1 in cells under mild oxidative stress (25  $\mu$ M H<sub>2</sub>O<sub>2</sub>) (Figure 4A), where Prdx1 was found to bind PTEN (Figure 3B). However, under higher oxidative stress treatment (500  $\mu$ M H<sub>2</sub>O<sub>2</sub>), which resulted in decreased binding of PTEN and Prdx1 (Figure 3B), impaired PTEN's lipid phosphatase activity. Western blotting confirmed equal amounts of PTEN protein in the lipid phosphatase assay (Figure 4A, lower panel). To complement those results, we exposed recombinant purified His-tagged PTEN to H<sub>2</sub>O<sub>2</sub> in the absence or presence

of increasing amounts of purified recombinant Prdx1 and assayed PTEN for PI(3,4,5)P<sub>3</sub> utilization. As expected (Figure 4B), in the presence of H<sub>2</sub>O<sub>2</sub>, we found that low molar amounts of Prdx1 protein weakly protected PTEN lipid phosphatase activity, whereas equimolar Prdx1 fully restored and further increases in Prdx1 protein amount did not enhance PTEN lipid phosphatase activity any more. Pre-incubation of PTEN in this assay with N- and C-terminal Prdx1 peptide replica (P1 and P2, respectively; peptide sequences were based on Prdx1 interaction mapping as shown in Supplementary Figure S3G and discussed for Figure 3G) decreased PTEN lipid phosphatase activity efficiently to 53%, which was comparable to PTEN activity observed with H<sub>2</sub>O<sub>2</sub> treatment in the absence of Prdx1 (50%) (Figure 4C). Addition of P1 decreased PTEN lipid phosphatase activity only to 65%, whereas P2 lowered it to 57%. However, as shown in Figure 4D, the addition of His-purified PTEN to recombinant Prdx1 inhibited Prdx1 peroxidase activity. Under our experimental conditions, the kinetics of NADPH





**Figure 3** (A) Epitope-tagged PTEN was expressed in 293T cells. Cell lysate was prepared under anaerobic conditions and immunoprecipitates were prepared as described in Materials and methods. A measure of 1000 μg of protein was immunoprecipitated (anti-PTEN antibody; Santa Cruz). Proteins were analysed by SDS-PAGE. PTEN and Prdx1 proteins were detected by staining membranes with anti-PTEN (138G6, Cell Signaling) and anti-Prdx1 (Abcam) antibodies, respectively. (B) Epitope-tagged PTEN and Prdx1 wild type were co-expressed in 293T cells. Before lysis, cells were treated with increasing dosages of H<sub>2</sub>O<sub>2</sub> as indicated for 15 min. Cell lysates were prepared under anaerobic conditions, and precipitated over night using HA-conjugated agarose beads. IPs were washed four times with argon-purged lysis buffer and analysed by western blotting. Proteins were detected with anti-PTEN, anti-Prdx1-SO<sub>2</sub>/SO<sub>3</sub> and anti-Prdx1 antibodies. Epitope-tagged Prdx1 migrates slower electrophoretically than endogenous Prdx1, labelled as HA-Prdx1. Epitope-tagged PTEN is labelled as Myc-PTEN. Anti-Prdx1-SO<sub>2</sub>/SO<sub>3</sub> can cross-react with Prdx2-4 and stains more intense in the IP proteins due to over night incubation and longer exposure to atmospheric oxygen. In contrast, lysates were immediately frozen (Supplementary Figure S3A and B). (C) Myc-PTEN and HA-Prdx1<sup>C51/172S</sup> were analysed as described under (B). Anti-Prdx1-SO<sub>2</sub>/SO<sub>3</sub> does not bind Prdx1<sup>C51/172S</sup>. (D) Recombinant His-PTEN (100 nM) was applied to equimolar GST or GST-Prdx1 and incubated over night. Glutathione sepharose (GST) beads were added for GST-pull down and analysed by western blotting. Input represents 1/50 of the total. (E) HA-Prdx1 wild type and cysteine to serine mutants of catalytically active Prdx1 cysteines were co-expressed with Myc-tagged PTEN wild type. HA-IPs were processed as under (B). Left side shows co-immunoprecipitations, right side shows expression of PTEN and Prdx1 cysteine to serine mutants. (F) Schematic domain structure of potential interaction sites of Prdx1 with PTEN (upper schematic) and PTEN with Prdx (lower schematic). (G) Identification of potential interaction sites using computer modelling (SwissPdb Viewer version 4; <http://www.expasy.org/spdbv>) (Guex and Peitsch, 1997). \*Prdx1 N terminus.



**Figure 4** (A) Epitope-tagged Myc-PTEN and HA-Prdx1 were co-expressed in 293T cells, which were treated with H<sub>2</sub>O<sub>2</sub> for 15 min, as indicated. Cell lysates were prepared under anaerobic conditions, and precipitated over night as described above by using anti-PTEN antibody. PTEN-immunoprecipitates were analysed for PI(3,4,5)P<sub>3</sub>-utilization in a 96-well plate assay as described in Materials and methods (triplicate samples). All results are presented as mean ± s.e. (or difference) for at least two independent experiments. Equal PTEN protein amount was confirmed by western blotting. (B) 500 μM of H<sub>2</sub>O<sub>2</sub> was added to either 20 pmoles of recombinant His-purified PTEN in the presence of different amounts of recombinant purified Prdx1 as indicated and described in Materials and methods. Experiment shown is representative of three independent studies. (C) PTEN activity was measured as described in (B) in the presence of peptides interfering with Prdx1 binding to PTEN. P1 (peptide 1) is replica of first N-terminal 21 amino acids (AS) of Prdx1; P1 Sc, scrambled sequence of P1; P2 (peptide 2), replica of last C-terminal 26 AS of Prdx1; P2 Sc, scrambled sequence of P2. The results are presented as mean ± s.e. (or difference) for at least two independent experiments. (D) Prdx1 peroxidase activity was measured by a standard thioresoxin (Trx)/thioredoxin reductase (TrxR)/NADPH-coupled spectrophotometric assay (see Materials and methods for details). A measure of 20 pmoles of purified Prdx1 (Sigma) was incubated with 500 μM H<sub>2</sub>O<sub>2</sub> and increasing molar amounts of purified His-PTEN as indicated. Prdx1 peroxidase activity (without PTEN) was about 68 nmol/mg protein/min under those conditions (extinction coefficient 6.62 mM<sup>-1</sup> cm<sup>-1</sup> for NADPH at 340 nm). Experiment shown is representative of three to four independent studies. (E) Relative reaction velocity of Prdx1 peroxidase activity from (D) was determined by plotting linear regression coefficient *K* ( $y = a - Kx$ , where *y* is the absorbency at 340 nm and *x* is time) versus various PTEN/Prdx1 (mol/mol) ratios. The results are presented as mean ± s.e. (or difference) for at least two independent experiments.

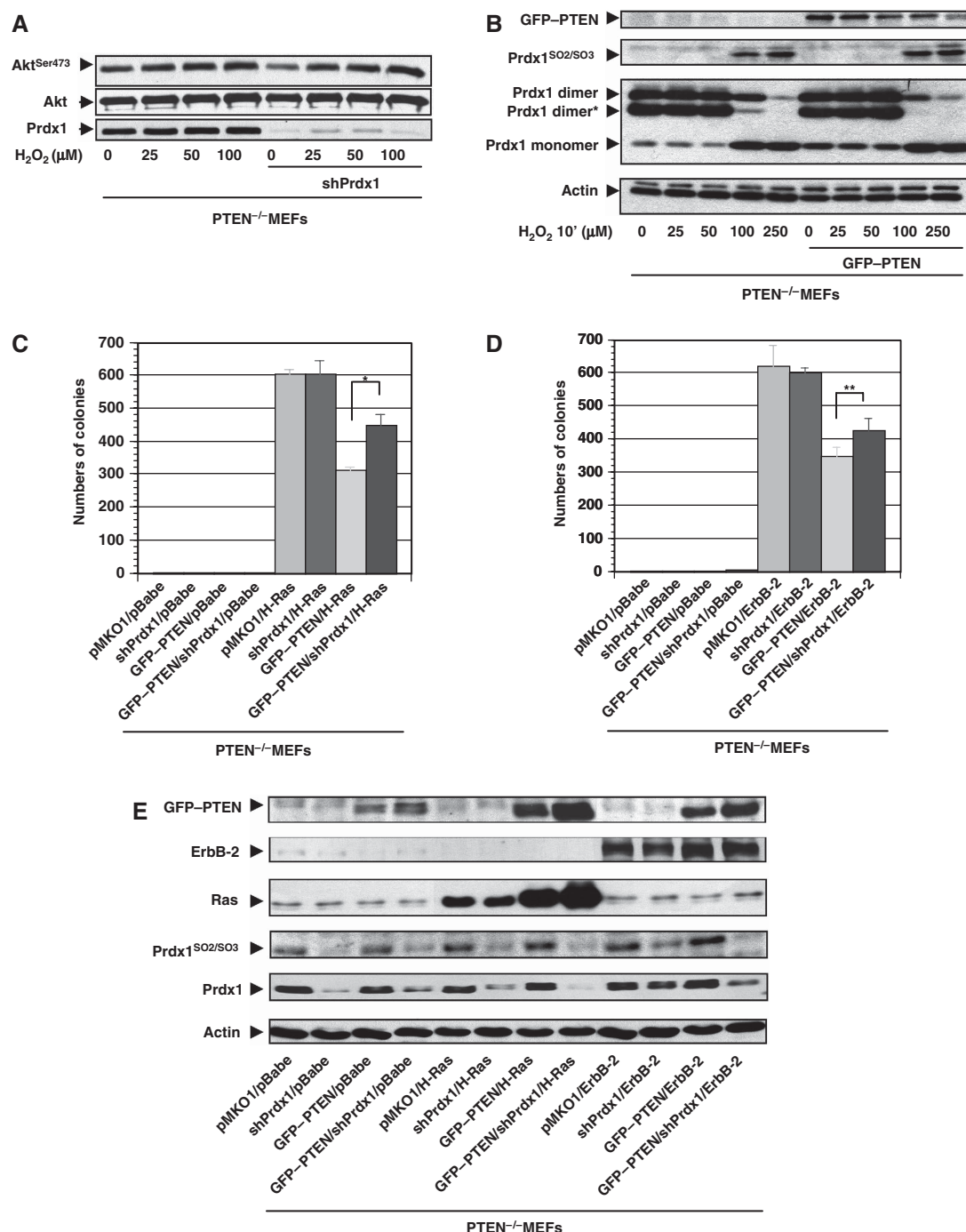
oxidation to NADP<sup>+</sup> were linear and the Prdx1 inhibition was proportional to molar amount of PTEN and reached saturation (completed inhibition) at PTEN/Prdx1=1:1 (mol/mol) ratio. Moreover, an increase in PTEN ratio lowered the velocity of Prdx1 peroxidase activity (Figure 4E).

#### Prdx1 suppresses H-Ras and ErbB-2-induced transformation mainly via promoting PTEN activity

Next, we examined in PTEN<sup>-/-</sup>MEFs whether Prdx1 has an important function in PTEN-induced tumour suppression. Although Prdx1 knock down in PTEN<sup>-/-</sup>MEFs decreased

pAkt<sup>Ser473</sup> levels, H<sub>2</sub>O<sub>2</sub> treatment of MEFs increased it equally in PTEN<sup>-/-</sup>MEFs and PTEN<sup>+/+</sup>MEFs (Figure 5A). Prdx1 is neither over-oxidized nor does it form less peroxidase active dimers in PTEN<sup>-/-</sup>MEFs compared with PTEN<sup>+/+</sup>MEFs reconstituted with GFP-PTEN, suggesting that in PTEN-null cells, Prdx1 activity is not compromised after H<sub>2</sub>O<sub>2</sub> treatment (Figure 5B). PI3K/Akt signalling is also essential for onco-

genic ErbB-2-induced transformation (Maroulakou *et al*, 2007), and Akt1 deficiency sufficiently suppresses tumour development in PTEN<sup>+/+</sup>mice (Chen *et al*, 2006). As shown for H-Ras (Figure 1B), Prdx1<sup>WT</sup> also prevented ErbB-2/neuT-induced transformation depending on its peroxidase activity (Supplementary Figure S5A). By using small hairpin (sh) RNA targeting Prdx1 mRNA (shPrdx1) in PTEN<sup>-/-</sup>MEFs,



**Figure 5** (A) PTEN<sup>-/-</sup>MEFs were serum starved for 48 h and treated with H<sub>2</sub>O<sub>2</sub> as indicated for 10 min. Protein lysates were analysed for pAkt<sup>Ser473</sup> as described before. (B) PTEN<sup>-/-</sup>MEFs were infected with retrovirus expressing GFP-PTEN. After 5-day selection in puromycin (2 μg/ml), MEFs were serum starved, exposed for 10 min to H<sub>2</sub>O<sub>2</sub> and analysed for Prdx1 oxidation and formation of dimeric structures by non-reducing SDS-PAGE. \*Prdx1 hetero-dimer formation with other Prdxs. (C) PTEN<sup>-/-</sup>MEFs were infected with retrovirus expressing shPrdx1, GFP-PTEN, H-Ras<sup>V12</sup> or various vectors carrying resistance gene only (pBabe, pMKO1). MEFs were plated in soft agar. Colonies were counted after 21 days. \**P* < 0.001 (Student's *t*-test). (D) As under C, except MEFs express ErbB-2/neuT instead of H-Ras<sup>V12</sup>. \*\**P* < 0.039 (Student's *t*-test). (E) Immunoblotting of MEFs from (C and D).



transformed by either H-RasV12 or ErbB-2/neuT, we showed that Prdx1 suppressed transformation mainly by regulating PTEN (Figure 5C–E). Although shPrdx1 expressed in PTEN<sup>-/-</sup>;H-RasV12 MEFs or PTEN<sup>-/-</sup>;ErbB-2/neuT MEFs did not increase colony formation compared with PTEN<sup>-/-</sup>;H-Ras;pMKO1 MEFs or PTEN<sup>-/-</sup>;ErbB-2/neuT;pMKO1 MEFs, in PTEN<sup>-/-</sup>;shPrdx1;GFP-PTEN,H-RasV12 MEFs showed 1.5-fold increase in colony formation compared with PTEN<sup>-/-</sup>;pMKO1;GFP-PTEN,H-RasV12 MEFs and PTEN<sup>-/-</sup>;shPrdx1;GFP-PTEN,ErbB-2/neuT MEFs and 1.35-fold increase in colony formation compared with PTEN<sup>-/-</sup>;pMKO1;GFP-PTEN;ErbB-2/neuT MEFs. Furthermore, a slight increase of Prdx1 oxidation was noted in PTEN<sup>-/-</sup>;pMKO1;H-RasV12 MEFs compared with PTEN<sup>-/-</sup>;GFP-PTEN;H-RasV12 MEFs and in PTEN<sup>-/-</sup>;pMKO1;ErbB-2/neuT MEFs compared with PTEN<sup>-/-</sup>;GFP-PTEN;ErbB-2/neuT MEFs. By using minimum dosages of two well-known PI3K inhibitors, Wortmannin (WM) and Ly294002 (Ly) that inhibited H<sub>2</sub>O<sub>2</sub>-induced Akt activity in Prdx1<sup>-/-</sup> MEFs less efficiently than in Prdx1<sup>+/+</sup> MEFs (Supplementary Figure S5B), we found that partial inhibition of PI3K/Akt activity translates into less efficient inhibition of H-Ras or ErbB-2-induced transformation in Prdx1<sup>-/-</sup> MEFs compared with Prdx1<sup>+/+</sup> MEFs (Table 1): 5 nM WM reduced Prdx1<sup>-/-</sup>;H-RasV12 MEFs colony number by <4%, 10 nM WM by 2.3%. However, 5 nM WM decreased Prdx1<sup>+/+</sup>;H-RasV12 MEFs colony numbers by 46% and ( $P = <0.001$ ) 10 nM WM by 67%. Similarly, 10  $\mu$ M Ly decreased Prdx1<sup>-/-</sup>;H-RasV12 MEFs colony number by 61% and in Prdx1<sup>+/+</sup>;H-RasV12 MEFs by 73%. However, 10  $\mu$ M Ly decreased colony number of Prdx1<sup>-/-</sup>;ErbB-2/neuT MEFs by only 53%, whereas in Prdx1<sup>+/+</sup>;ErbB-2/neuT MEFs by 75%. Moreover, 25  $\mu$ M Ly decreased colony formation equally in Prdx1<sup>-/-</sup>;ErbB-2/neuT MEFs and Prdx1<sup>+/+</sup>;ErbB-2/neuT MEFs to 4–5%.

#### Loss of Prdx1 promotes H-Ras-induced mammary tumours and PTEN oxidation in mammary epithelial cells

MMTV/v-H-Ras mice transgenic mice are highly susceptible to mammary carcinomas due to the targeted over-expression of the activated Ras oncogene in their mammary glands (Sinn *et al*, 1987) and Akt1 ablation lowers the incidence of tumours in these mice (Maroulakou *et al*, 2007). In our experiment, we confirmed that Prdx1 plays a role in Ras-induced tumorigenesis *in vivo* (Figure 6A). In all, 60% of the Ras/Prdx1<sup>+/+</sup> mice remained tumour free during the entire

observation period of 36 weeks. In contrast, only 40% of the Ras/Prdx1<sup>+/-</sup> mice remained tumour free during that time with a median survival ( $T_{50}$ ) of 28 weeks, which indicated that the loss of one copy of Prdx1 accelerated Ras-induced breast cancer. Moreover, eliminating both copies of Prdx1 in MMTV/v-H-Ras mice (Ras/Prdx1<sup>-/-</sup>), the  $T_{50}$  for breast cancer was only 20 weeks. We then evaluated basal PTEN oxidation and pAkt<sup>Ser473</sup> in mammary epithelial cells (MECs) from 16-week-old mice that had not developed tumours. Lack of Prdx1 increased basal PTEN oxidation (Figure 6B) and pAkt<sup>Ser473</sup> (Figure 6C).

## Discussion

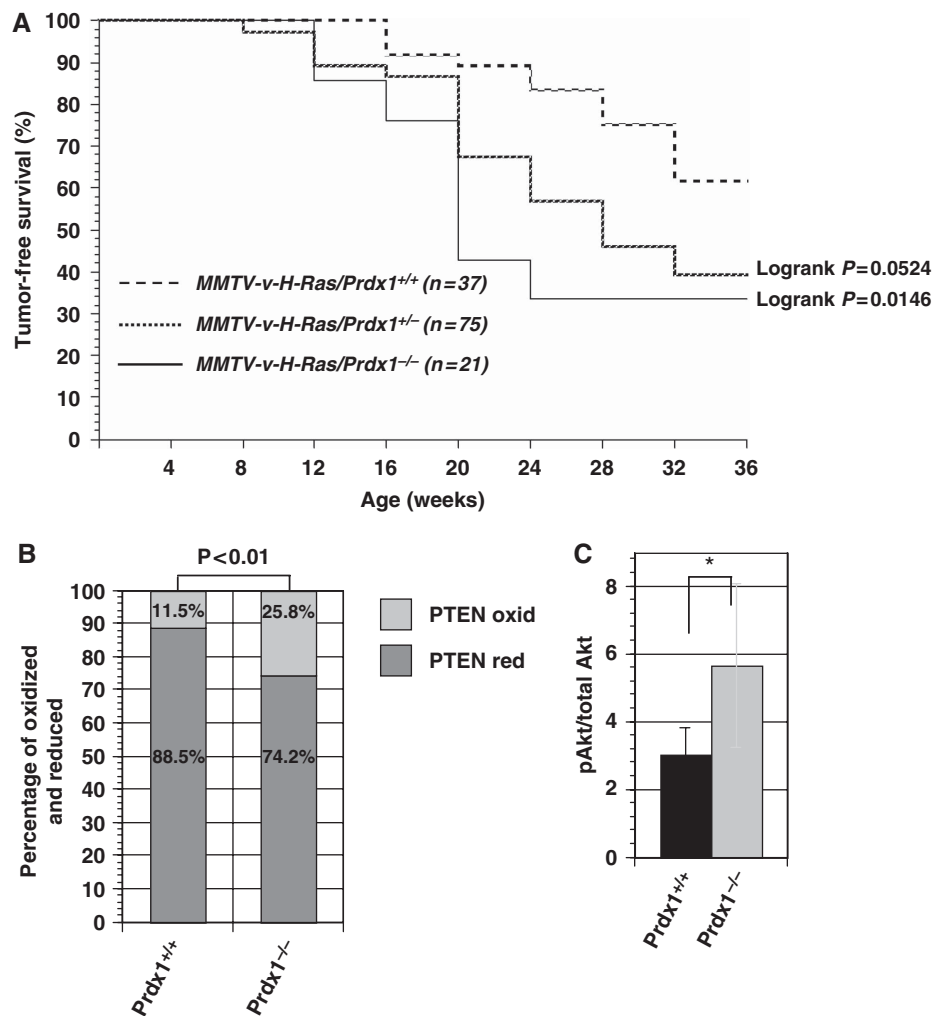
Compared with Prdxs, catalase peroxidase activity decomposes H<sub>2</sub>O<sub>2</sub> with a >1000-fold higher  $K_M$  (Chae *et al*, 1994a; Loewen *et al*, 2004) and therefore eliminates H<sub>2</sub>O<sub>2</sub> with much higher efficiency. Thus, we cannot expect that Prdx1 scavenges H<sub>2</sub>O<sub>2</sub> in considerable amounts, as shown in Figure 1A and Supplementary Figure S1A. Yet, Prdx1's peroxidase activity is important in preventing H-Ras<sup>V12</sup> (Figure 1B) or ErbB-2/neuT (Supplementary Figure S5A) induced transformation. Similar to the catalase knockout mice, all other currently published Prdxs knockout mice, show phenotypically no increased susceptibility to transformation (Ho *et al*, 1997). Therefore, Prdx1 may have a unique function amongst cellular peroxidases in tumour suppression, by affecting signalling directly through physical interaction with target enzymes. This was shown for c-Abl tyrosine kinase (Wen and VanEtten, 1997) and c-Jun terminal kinase (Kim *et al*, 2006). As described here for PTEN (Figure 3B), oxidative stress induced Prdx1 to dissociate, which in case of c-Abl (Neumann *et al*, 1998) and JNK (Kim *et al*, 2006) resulted in kinase activation. The Prdx1:PTEN heterodimer is most likely formed in a 1:1 molar ratio, as (1) the Prdx1 preservation of PTEN lipid phosphatase activity under oxidative stress is achieved by a 1:1 (mol:mol) ratio of Prdx1 and PTEN and could not be further increased by excess of Prdx1 (Figure 4B) and (2) Prdx1<sup>C51/172S</sup>, which is unable to form dimeric structures (Chae *et al*, 1994b), bound to PTEN and did not dissociate under H<sub>2</sub>O<sub>2</sub> treatment (Figure 3C). Therefore, these data provide compelling evidence that Prdx1 may bind PTEN as a monomer.

We further propose that the Prdx1:PTEN interaction is essential for protecting PTEN from oxidation-induced inactivation (Figure 4C), as Prdx1 C- and N-terminal peptide

**Table 1** Prdx1<sup>-/-</sup> MEFs have a higher sensitivity to PI3K inhibitors in H-Ras or ErbB-2 induced transformation

Wortmannin (nm)		H-Ras colony number in %	Ly294002 ( $\mu$ M)		H-Ras colony number in %	ErbB-2/neuT colony number in %
0	Prdx1 <sup>-/-</sup>	100 $\pm$ 23.3	0	Prdx1 <sup>-/-</sup>	100 $\pm$ 11.2	100 $\pm$ 4.6
5		96.3 $\pm$ 5.7	10		38.3 $\pm$ 15.2	47.3 $\pm$ 6.6
10		97.8 $\pm$ 13.5	25		5 $\pm$ 10.1	3.2 $\pm$ 4.1
0	Prdx1 <sup>+/+</sup>	100 $\pm$ 4.1	0	Prdx1 <sup>+/+</sup>	100 $\pm$ 11.2	100 $\pm$ 4.6
5		55.9 $\pm$ 12.5	10		26.3 $\pm$ 5.5	25.1 $\pm$ 6.6
10		32.7 $\pm$ 24.4	25		4 $\pm$ 8.7	3.2 $\pm$ 4.8

Prdx1<sup>-/-</sup> MEFs and Prdx1<sup>+/+</sup> MEFs were retrovirally infected with constructs carrying genes for H-Ras<sup>V12</sup> or ErbB-2/neuT (ErbB-2Val644Glu) and treated after plating in soft agar either with 10 or 25  $\mu$ M Ly294002 every 2 days or Wortmannin 5 or 10 nM every day. Colonies were counted after 21 days. Decrease in colony formation (%) relates to untreated of the same genotype (%). This experiment is representative of three independent studies with two different sets of MEF clones from Prdx1 littermates.



**Figure 6** (A) The percentage of mice free of H-Ras-induced mammary tumours is shown in MMTV-v-H-Ras mice carrying only one, none, or both copies of the *Prdx1* gene. (B) MECs ( $n=3$ ) were isolated from MMTV-v-H-Ras mice/*Prdx1*<sup>-/-</sup> or MMTV-v-H-Ras mice/*Prdx1*<sup>+/+</sup> and analysed on a non-reducing SDS-PAGE for PTEN oxidation as described above. Statistical analysis of PTEN oxidation was done using a  $\chi^2$  test. (C) The same protein lysates were also analysed for Akt phosphorylation on Ser473, as described above. \*Non-statistically significant.

replica, which were designed based on our mapping studies and computer modelling to block PTEN docking sites on Prdx1 (Figure 3F and G; Supplementary Figure S3E), resulted after H<sub>2</sub>O<sub>2</sub> treatment in a 47% reduction of PTEN activity. This was comparable to PTEN activity after H<sub>2</sub>O<sub>2</sub> treatment in the absence of Prdx1 (50%) (Figure 4C). The Prdx1 N terminus, which includes Cys51, may have an important function in regulating the H<sub>2</sub>O<sub>2</sub>-induced dissociation of the Prdx1:PTEN complex, as Cys51 when replaced by a serine enhances Prdx1 binding to PTEN (Figure 3D). X-ray crystallography of Prdxs suggested significant conformational changes, such as unwinding of the active site N-terminal helix, to form disulfides, as in reduced Prdxs, the sulfur atoms of the N- and C-terminal conserved cysteine residues are too far apart to react with each other (Schroder *et al*, 2000; Matsumura *et al*, 2008). Therefore, in the Prdx1:PTEN complex, increased H<sub>2</sub>O<sub>2</sub> may oxidize Cys51 (Prdx1), thus, promoting unwinding of the PTEN-binding conformation of Prdx1 consequently inducing dissociation.

Over-oxidation of Prdx1 Cys51 occurs during catalysis and correlates with increasing amounts of Trx (Yang *et al*, 2002). The reduction of Cys51 by Trx inhibits the folding back of Cys51 into its pocket thereby exposing it to further oxidation

by H<sub>2</sub>O<sub>2</sub>. The binding of PTEN to Prdx1 could have similar effects on Prdx1, thereby inhibiting sufficient reduction of Cys51 by Trx and decreasing Prdx1 peroxidase activity (Figure 4C and D). Alternatively, as it has been shown that Trx binds PTEN and inactivates its lipid phosphatase activity, it could be that under our experimental conditions Trx is depleted from the Prdx1 scavenging system by binding PTEN (Meuillet *et al*, 2004). This seems unlikely though, as Trx was added in approximately 300-fold molar excess, which should provide a large Trx pool to reduce oxidized Prdx1 proteins.

We defined a novel interaction site of Prdx1 with PTEN in the C2 domain of PTEN (Figure 3F; Supplementary Figure S3D and E), which together with the N-terminal PTD, are both required for enzyme activity. Surface plasmon resonance analysis revealed that the C2 domain is essential for high-affinity membrane binding of PTEN (Das *et al*, 2003). Our data support this finding, as it has been shown recently through chemical proteomic strategy using cleavable lipid baits, that Prdx1 is a potential novel phosphoinositides-binding protein (Pasquali *et al*, 2007). Prdx1 may, therefore, stabilize PTEN binding to the plasma membrane, which in a recent analysis using single-molecule TIRF microscopy,

lasts only for a few hundred milliseconds (Vazquez *et al*, 2006). Such short time of membrane binding could explain why we (Figure 2B, C, E and F; Supplementary Figure S2B and D) and others see only small amounts of PTEN oxidized after cell exposure to H<sub>2</sub>O<sub>2</sub> or PDGF (Kwon *et al*, 2004). PDGF-induced H<sub>2</sub>O<sub>2</sub> stems either from metabolized superoxide released by NADPH oxidases (NOXs) or the mitochondria and is considered a signalling molecule (Rhee, 2006). As discussed earlier (Leslie *et al*, 2003), such finding may be attributed to the proximity of NOXs complexes and PIP<sub>3</sub>. Over-expression of Nox1, in PDGF-stimulated NIH 3T3 cells rapidly increases PIP<sub>3</sub> levels, in contrast to PI3K activity, suggesting that Nox1-induced superoxide enhances PIP<sub>3</sub> levels by inactivating PTEN rather than activating PI3K. Expression of a peroxidase inactive Prdx2 leads to a rapid increase of PIP<sub>3</sub> levels in growth factor stimulated cells, which is reversed by Prdx2<sup>WT</sup> (Kwon *et al*, 2004). In our hands however, Prdx2 did not bind PTEN, as we were unable to detect an interaction of Prdx2 and PTEN by immunoprecipitation (data not shown).

PTEN lipid phosphatase activity has an important function in tumour suppression (Tolkacheva and Chan, 2000; Koul *et al*, 2002) and Akt1 ablation protects (MMTV)-ErbB2/neu, MMTV-polyoma middle T and MMTV-v-H-Ras mice from breast cancer initiation (Skeen *et al*, 2006; Maroulakou *et al*, 2007). Here, we show that loss of Prdx1 increased basal, H<sub>2</sub>O<sub>2</sub> and PDGF-induced Akt activity (Figure 2A, D and G) and Prdx1 peroxidase activity was essential in suppressing H-RasV12 and ErbB-2/neuT-induced transformation (Figure 1B; Supplementary Figure S5A). Along those lines, Prdx1 ablation accelerated mammary tumorigenesis in MMTV-v-H-Ras mice and increased levels of pAkt<sup>Ser473</sup> and PTEN oxidation in MECs from these mice (Figure 6A–C). Prdx1 tumour suppression seemed to occur largely via regulation of the PTEN and Akt. Knock down of Prdx1 in PTEN<sup>−/−</sup>MEFs did not significantly enhance H-Ras or ErbB-2-induced transformation (Figure 5C and D), whereas in transformed PTEN-containing MEFs, decrease in Prdx1 levels resulted in a gain of transformation: 1.5-fold for H-RasV12 and only 1.35-fold for ErbB-2/neuT, which we contributed to decreased Prdx1 knock down in PTEN<sup>−/−</sup>;shPrdx1;GFP-PTEN;ErbB-2/neuT MEFs compared with PTEN<sup>−/−</sup>;shPrdx1;GFP-PTEN;H-RasV12 MEFs. Lack of PTEN also did not result in Prdx1 over-oxidation nor reduced expression (Figure 5B and E), suggesting that in PTEN-deficient cells, Prdx1 is acting as peroxidase.

However, we have to consider the regulation of other phosphatases or kinases by Prdx1 or H<sub>2</sub>O<sub>2</sub> in Akt-driven tumorigenesis. SHIP-1 and 2 are known to dephosphorylate PIP<sub>3</sub> to PtdIns (3,4)P<sub>2</sub> (Vivanco and Sawyers, 2002). However, SHIP2 is not inactivated by oxidative stress (Leslie *et al*, 2003). PTEN<sup>−/−</sup>;shPrdx1 MEFs showed decreased pAkt<sup>Ser473</sup> levels compared with PTEN<sup>−/−</sup>MEFs (Figure 5D), perhaps due to the activation of SHIP2 by small amounts of H<sub>2</sub>O<sub>2</sub>. However, H<sub>2</sub>O<sub>2</sub> treatment of PTEN<sup>−/−</sup>;shPrdx1 MEFs increased pAkt<sup>Ser473</sup> levels comparable to PTEN<sup>−/−</sup>MEFs, suggesting maybe a H<sub>2</sub>O<sub>2</sub>-induced inactivation of PP2A (Rao and Clayton, 2002), as loss of Prdx1 also inactivated PP2A activity (Supplementary Figure 2E). We also cannot exclude a regulation of PI3K by Prdx1, as H-Ras and ErbB-2/neuT transformed Prdx1<sup>−/−</sup>MEFs display a lesser sensitivity

towards PI3K inhibitors (Table I; Supplementary Figure 5B), compared with Prdx1<sup>+/+</sup>MEFs.

In summary, oxidative stress and carcinogenesis are closely linked, however, molecular details, especially how oxidative stress promotes pro-oncogenic signalling pathways are scarce. Our studies shed some light on how H<sub>2</sub>O<sub>2</sub> can promote oncogenic Ras and ErbB-2 signalling by providing evidence for the first time that the tumour suppressive function of PTEN and Prdx1 are closely related. The mechanistic details of the Prdx1 and PTEN interaction and its regulation in terms of Prdx1 peroxidase activity and PTEN oxidation will be important in future work.

## Materials and methods

### Reagents

All chemicals were purchased from Sigma Aldrich unless otherwise indicated. Ly294002, WM, antibodies against PTEN (138G6), Akt (11-E7), Akt-phosphoserine 473 (193H12), Akt-phosphothreonine 308 (244F9), phospho Akt substrate (110B7E) and HA-tag were purchased from Cell Signaling; Antibodies recognizing PTEN (A2B1), PTEN (N-19) and Actin (C-11) were purchased from Santa Cruz; antibodies against Prdx1 and Prdx1-SO<sub>2</sub>/SO<sub>3</sub> were purchased from Abcam; Ras antibody from Oncogene; ErbB-2 antibody from Biosource. HA-conjugated agarose beads were purchased from Roche. Amplex Red reagent: Molecular Probes. DMEM, FBS, Glutamax, NEAA, Pen/Strep, Sodium Pyruvate, DMEM without Phenol Red, PBS and PDGF stem from Invitrogen; GST-sepharose from Pharmacia. 6His-PTEN was expressed in *Escherichia coli* BL21 from the fusion vector pProEx HTb PTEN and purified by standard protocols.

### Retrovirus

The retrovirus was generated after conducting a transient transfection of retroviral constructs into a 293T/17 (ATCC) ecotropic-packaging cell line. Retroviral infection of MEFs was carried out for 4–6 h in the presence of 8 µg/ml polybrene. An antibiotic selection was carried out 24 h later until all uninfected cells were eliminated and stable polyclonal cell lines were generated.

### Generation of MEFs

MEFs were generated from Prdx1<sup>+/−</sup> littermate matings in a C57BL/6J/129Sv background. Primary MEFs were harvested from E13.5 embryos as described earlier (Neumann *et al*, 2003). Primary Prdx1 and PTEN MEFs (passages 3–4) were immortalized by retroviral infection with pBabe-hygro DD p53 expressing dominant-negative p53 (Hahn *et al*, 2002) and selected with 150 µg/ml hygromycin for 14 days.

### Amplex red assay

In total, 35 000 MEFs were plated (*n* = 6) in 24-well plates (Costar 24) coated with fibronectin. On the day of the assay, cells were washed with PBS and replated with serum-free and phenol red-free DMEM medium. Amplex Red reagent and horse radish peroxidase (HRP) were prepared as recommended by the manufacturer (HRP 0.2 U/ml). MEFs were incubated with Amplex Red/HRP solution at 37°C before the first reading (T<sub>0</sub>) at 540 nm on a plate reader (Molecular Devices SpectraMax M5). Plates were kept in an incubator between readings. The resulting fluorescence (FLU) was compared with FLU from H<sub>2</sub>O<sub>2</sub> standard curves and analysed.

### Soft agar assays

P53DD-immortalized MEFs (Prdx1<sup>−/−</sup>, Prdx1<sup>+/+</sup>) were infected with retrovirus containing either Prdx1<sup>WT</sup> or Prdx1<sup>C172/51S</sup>, both subcloned into pQCXI-puro (Clontech) and selected 10 days in puromycin 2 µg/ml. Some clones were further infected with a retrovirus containing either H-Ras<sup>V12</sup> (pBabe-puro H-Ras<sup>V12</sup>) (Yu *et al*, 2001) or wild-type ErbB-2 (pBabe-puro ErbB-2) (Yu *et al*, 2001) and selected in 5 µg/ml puromycin for another 5 days. MEFs were then resuspended in DMEM containing 0.53% agarose (Difco) and plated onto a layer of 0.7% agarose-containing medium in 3.5-cm plates in duplicate (10 000 cells per plate). Colonies were counted after 14–21 days.

### Akt activity and PTEN oxidation

MEFs ( $1.8 \times 10^5$ ) were plated on 10-cm plates and serum-starved (0.25% FBS) for 48 h. Plates were then incubated with  $H_2O_2$  ( $\pm$  Ly294002 or WM) as indicated. Akt and PTEN protein analysis were done as described earlier (Leslie *et al*, 2003) with slight modifications. Briefly, lysis buffer (50 mM Tris pH 7.4; 1% Triton 100; 0.5 mM EDTA; 0.5 mM EGTA; 150 mM NaCl; 10% Glycerol; 50 mM NaF; 100 mM  $NaVO_4$ ; 40 mM  $\beta$ -Glycerophosphate) was degassed with Argon for 20 min before adding 1% TritonX-100; 2 mM PMSF; 5 mg/ml Aprotinin; N-ethylmaleimide 40 mM; and bovine catalase 100  $\mu$ g/ml. Plates were transferred into an anaerobic/argon cabinet and were washed twice with cold degassed PBS. Cells were then scraped into lysis buffer and lysed on ice for 20 min before conducting quantitative analysis as described above. A measure of 65  $\mu$ g of protein were analysed under non-reducing conditions with a 7.5% SDS-PAGE.

### Co-immunoprecipitation

PTEN<sup>WT</sup> (Addgene) was subcloned into a pQCXI vector (Clontech) and N-terminally tagged with Myc epitope. Prdx1<sup>WT</sup> and Prdx1<sup>C51/172S</sup>, Prdx1<sup>C51S</sup>, Prdx1<sup>C172S</sup> (generated by site-specific mutagenesis) were subcloned into a pCGN vector and N-terminally tagged with an HA epitope. All constructs were transfected into 293T/17 (ATCC) cells using Fugene transfection reagent (Roche). Cells were harvested 72 h later for protein lysis under anaerobic conditions (as described above). A measure of 1000  $\mu$ g protein was immunoprecipitated using anti-HA affinity matrix overnight at 4°C before washing four times with degassed lysis buffer before analysis on 4–12% gradient SDS-PAGE. Proteins were detected as described above.

### GST-pull down

BL21 (DE3) were transformed and cultured overnight with pGEX-4T-2 (Amersham) ligated earlier with Prdx1. At absorbance of 0.6–0.8 at 600 nm, GST-Prdx1 expression was induced by the addition of IPTG (200  $\mu$ M). Bacterial pellets were re-suspended using fusion protein buffer containing 140 mM NaCl, 10 mM  $Na_2HPO_4$ , 1.8 mM  $KH_2PO_4$ , 2.7 mM KCl, 1 mM DTT, 50  $\mu$ M PMSF, 5 mM benzamidine hydrochloride hydrate and 3  $\mu$ M aprotinin. Lysate was then sonicated and proteins were solubilized by the addition of Triton X-100 (2%). After incubation of supernatant with glutathione affinity matrix, the matrix was washed consecutively with 500 mM NaCl, 50 mM NaCl and the GST-tagged fusion protein was eluted from the matrix by incubation with 30 mM reduced glutathione. The fusion protein was then desalted into 20 mM Tris (pH 7.4) and concentrated by centrifugation using a Centricon 30 kD MWC filter (Millipore, Bedford, MA). Incubations of equimolar concentrations (100 nM) of GST-Prdx1 plus His-PTEN or GST-EV (empty vector) plus His-PTEN were performed in a buffer containing 20 mM Hepes (pH 8.0), 150 mM NaCl, 2 mM  $MgCl_2$ , 1 mM DTT, 50  $\mu$ M PMSF, 5 mM benzamidine hydrochloride, 3  $\mu$ M aprotinin and 1% Triton X-100. Mixtures were allowed to incubate overnight at 4°C before pre-washed glutathione agarose beads were added. The retained proteins were eluted from the resin by addition of 35  $\mu$ l of 4  $\times$  loading buffer and analysed by 10% SDS gels.

### PTEN Plasmids

Different lengths of cDNA fragments encoding human PTEN amino acids from 1–403 were subcloned from pSG5L HA-PTEN<sup>WT</sup> (Addgene#10750), pSG5L-HA-PTEN<sup>C124S</sup> (#10745), pSG5L-HA-PTEN<sup>1–274</sup> (#10740), pSG5L-HA-PTEN<sup>1–373</sup> (#10766), pSG5L-HA-PTEN <sup>$\Delta$ 89–138</sup> (#10924), pGEX2T-PTEN <sup>$\Delta$ 274–342</sup> (#10739) (Ramswamy *et al*, 1999) at restriction sites *Bam*HI/*Eco*RI. A double-stranded oligonucleotide encoding the Myc-tag sequence was inserted 5' of the PTEN to generate Myc-PTEN. Inserts tagged with Myc tag were cloned into pQCXIP to generate the corresponding pQCXIP-Myc-PTEN plasmids; pQCXIP-MYC-PTEN<sup>1–185</sup> or PTEN<sup>1–351</sup> or PTEN<sup>186–351</sup> or PTEN <sup>$\Delta$ 244–251</sup> were generated by PCR mutagenesis and confirmed by DNA sequencing. Prdx1 N- and C-terminal truncation mutants were generated by Bal31 nuclease digest, fill-in with T4-polymerase and subcloned into pCGN-HA.

### Recombinant Prdx1 activity assay

Prdx1 peroxidase activity was studied by standard Trx/Trx reductase (TrxR)/NADPH-coupled spectrophotometric assay as described (Chae *et al*, 1994b). Briefly, NADPH oxidation coupled to the reduction of  $H_2O_2$  was monitored at 37°C as a decrease in

A<sub>340</sub> using a SpectraMax M5 plate reading spectrophotometer. The reaction was initiated by addition of the  $H_2O_2$  (500  $\mu$ M) as a substrate into a reaction mixture containing 50 mM Hepes (pH 7.0), 0.15 U TrxR, 18  $\mu$ M Trx, 25  $\mu$ M NADPH and 680 nM Prdx1 (Sigma). The effect of PTEN on Prdx1 activity was studied by titration of reaction mixture with indicated increasing amount of PTEN (0.25–1.25 PTEN/Prdx1 mol/mol ratio). All experimental data were normalized to initial (before addition of substrate) absorbency and fitted with linear regression (Sigma Plot 10, Systat Software, Inc., San Jose, CA). Quality of fit was controlled by regression coefficient ( $R^2 \geq 0.9$ ).

### PTEN inositol phosphatase activity

Myc-PTEN was expressed in 293T/17 cells in the presence or absence of HA-Prdx1. Cells were lysed as described above in phosphatase inhibitors free lysis buffer. PTEN proteins were immunoprecipitated from lysates with anti-PTEN (Santa Cruz) o/n as described above. IPs were then washed twice as described (Schwartzbauer and Robbins, 2001), followed by a final wash in PTEN enzyme reaction buffer (10 mM Hepes, 150 mM NaCl, 10 mM DTT pH 7.2). The phosphatase reactions were done following manufacturer's instructions (Echelon Biosciences). Briefly, the immunoprecipitated PTEN proteins were incubated with PTEN enzyme reaction buffer for an appropriate amount of time at 37°C in a 96-well plate coated with PI(3,4,5)P<sub>3</sub>. After removal of PTEN proteins from plates, PI(4,5)P<sub>2</sub> was determined according to the manufacturer's instructions. Absorbance was measured at 450 nm in a microplate reader, and PI(4,5)P<sub>2</sub> produced was calculated from a standard curve.

### Recombinant PTEN inositol phosphatase activity

A measure of 20 pmol purified recombinant human His-PTEN was incubated with different amounts of Prdx1 (Sigma Aldrich) and 500  $\mu$ M  $H_2O_2$  in 20 mM Tris-Cl (pH 7.4) buffer at room temperature for 10 min.  $H_2O_2$  was removed by Bio-Gel P-6 chromatography columns (Bio-Rad), following the manufacture's instructions. The protein mixture was then loaded onto PTEN activity assay plates as described above (Echelon Biosciences). Equal amount of PTEN was confirmed by western blot with the PTEN antibody. For Prdx1 peptide interference, 20 pmoles purified recombinant human His-PTEN was pre-incubated with 20-fold excess amount of peptides on ice in 20 mM Tris-Cl (pH 7.4) (for Peptide 1 or scramble peptide 1) or 0.2 mM Tris-Cl (pH 7.4) (peptide 2 or scramble peptide 2) for 1 h, the unbound peptides were removed by Microcon Ultracel YM-10 filters following the manufacture's instructions. The recovered PTEN-peptide mixtures were then incubated with 20 pmoles Prdx1 (Sigma Aldrich) and 500  $\mu$ M  $H_2O_2$  in 20 mM Tris-Cl (pH 7.4) buffer at room temperature for 10 min and further processed as described above. Peptide 1: MSSGNKIGHAPNFKATAVM; scrambled peptide 1: KTHPMPSIAAKFNAGSNAMVG; Peptide 2: PAGWKPGSDTIKPDVQK SKEYFSKQK; scrambled peptide 2: IKKSWKFGPGQPTSAPKQSKEV-KYDD; all peptides had purity >75% and were purchased from Genscript, NJ.

### PTEN<sup>-/-</sup> MEFs experiments

An optimal sequence for knock down of Prdx1 by sh (5'-GCTCAGGATTATGGAGTCTTA-3') was identified from the TRC consortium (<http://www.broad.mit.edu>) and was subcloned into pMKO1 retroviral vector by *Age*I/*Eco*RI digest. PTEN<sup>-/-</sup>MEFs were infected with retroviral supernatant containing either pMKO1-Prdx1shRNA or pMKO1 EV, and selected in puromycin 2  $\mu$ g/ml for 1 week. GFP-PTEN was subcloned from pEGFP-C2-PTEN (Leslie *et al*, 2000) into pQCXIP at restriction sites of *Age*I-*Eco*RI. PTEN<sup>-/-</sup>:pMKO1<sup>+</sup>MEFs or PTEN<sup>-/-</sup>:shPrdx1<sup>+</sup>MEFs were then transduced with GFP-PTEN retrovirus and selected in 6  $\mu$ g/ml puromycin. Infection and selection with retrovirus expressing either H-Ras<sup>V12</sup> or ErbB-2/neuT was then challenged by 10  $\mu$ g/ml puromycin.

### MMTV-v-H-Ras mice lacking Prdx1

MMTV-v-H-Ras mice (Sinn *et al*, 1987) were purchased from Charles River. Prdx1<sup>+/-</sup>/MMTV-v-H-Ras mice were intercrossed to generate Prdx1<sup>-/-</sup>/MMTV-v-H-Ras and Prdx1<sup>+/-</sup>/MMTV-v-H-Ras mice. Mice were monitored twice a week for tumour appearance. Mouse mammary glands were isolated from age-matched 16-week-old female MMTV-v-H-ras/Prdx1<sup>-/-</sup> and MMTV-v-H-ras/Prdx1<sup>+/-</sup> ( $n = 3$ ) mice following earlier described procedures (Fujiwara *et al*, 2005). Proteins were processed as described above.

## Supplementary data

Supplementary data are available at *The EMBO Journal* Online (<http://www.embojournal.org>).

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